

Supporting Information

Monitoring Solid-Phase Glycoside Synthesis with ^{19}F NMR Spectroscopy

Mickael Mogemark, Mikael Elofsson and Jan Kihlberg*

Organic Chemistry, Department of Chemistry, Umeå University, SE-901 87 Umeå, Sweden

General methods and materials. – All reactions were carried out in room temperature under an inert nitrogen atmosphere using dry, freshly distilled solvents under anhydrous conditions, unless otherwise stated. MeCN and CH_2Cl_2 were distilled from calcium hydride. Diethyl ether and THF were distilled from sodium benzophenone. MeOH and pyridine were dried over 3 Å and 4 Å molecular sieves, respectively. Organic solutions were dried over Na_2SO_4 before being concentrated. Solid phase synthesis was performed on ArgoGel- NH_2 resin (151 μm , loading capacity: 0.38 mmol/g) using a semi - automatic Quest 210 synthesizer, which agitates the resin by moving the magnetic stirring bar vertically in the reactor with an external magnet.

TLC was performed on Silica Gel F₂₅₄ (Merck) and detection was carried out by examination under UV light and by charring with 10% sulfuric acid. Flash column chromatography was performed on Silica Gel (Matrex, 60 Å, 35 – 70 μm , Grace Amicon). Preparative HPLC separations were performed on a Beckman System Gold

HPLC, using a Kromasil C-8 column (250×20 mm, 5 μ m, 100 Å) with a flowrate of 11 mL/min, detection at 214 nm, and the following eluent systems: *A*, aq. 0.1% CF₃CO₂H; and *B*, 0.1% CF₃CO₂H in MeCN. Analytical HPLC were performed on a Beckman System Gold HPLC, using a Kromasil C-8 column (250×4.6 mm, 5 μ m, 100 Å) with a flowrate of 1.5 mL/min, detection at 214 nm, and the following eluent systems: *A*, aq. 0.1% CF₃CO₂H; and *B*, 0.1% CF₃CO₂H in MeCN.

¹H and ¹³C NMR spectra were recorded with a Bruker DRX-400 spectrometer for solutions in CDCl₃ [residual CHCl₃ (δ_{H} 7.26 ppm), CDCl₃ (δ_{C} 77.0 ppm) as internal standard] or CD₃OD [residual CD₂HOD (δ_{H} 3.35 ppm), CD₃OD (δ_{C} 49.0 ppm) as internal standard] at 300 K. First order chemical shifts and coupling constants were determined from one-dimensional spectra and proton resonances were assigned from COSY and HETCOR experiments. Proton resonances that could not be assigned are not reported. Proton decoupled gel phase ¹⁹F NMR spectra were recorded with a Bruker DRX-400 spectrometer for resin suspensions in CDCl₃ [CFCl₃ (δ_{F} 0.00 ppm) as internal standard] at 300 K. Two peaks appear in the spectra around 0.00 ppm, one resonance originates from CFCl₃ inside the polymer the other resonance from CFCl₃ outside the polymer. The peak with highest shift was used as internal standard.

Resin 3

HOBt (185 mg, 1.37 mmol) and DIC (138 μ l, 0.89 mmol) were added to a solution of 3-fluoro-4-hydroxybenzoic acid **2** (142 mg, 0.91 mmol) in DMF (4 mL). The mixture was stirred for 20 min and then transferred to resin **1** (0.228 mmol) that had been preswollen in DMF and briefly washed with 20% piperidine in DMF (3 mL, 1 min agitation) and with DMF (5×3 mL). Bromophenol Blue (0.001 equiv) was added as

indicator of free amino groups and the mixture turned blue. After 12 h of vertical agitation the color of the reaction turned yellow, indicating that the reaction had reached completion. The solution was removed by filtration. The resin was washed with DMF (3×3 mL), 20% piperidine in DMF (3 mL, 1 min agitation), DMF (3×3 mL), and CH₂Cl₂ (5×3 mL). A solution of CH₂Cl₂ (5 mL) and NaOMe in MeOH (2 M, 0.4 mL) were added to the resin and after 2 h agitation the resin was washed with CH₂Cl₂, MeOH, CH₂Cl₂ containing 3% HOAc, CH₂Cl₂, DMF and CH₂Cl₂ (3×5 mL each). Resin 3 had: ¹⁹F NMR data (CDCl₃): δ -137.9 (s, 1F).

4-Hydroxybut-1-yl 4-fluorobenzoate (4)

4-Fluorobenzoyl chloride (1.18 mL, 10.0 mmol) was added dropwise over 10 min to a solution of 1,4-butanediol (5.32 mL, 60 mmol) in pyridine (30 mL). After stirring for 18 h the mixture was concentrated and residual pyridine was removed by co-evaporation with toluene. The residue was chromatographed (heptane/EtOAc 5:1 → 2:1) to give **4** (1.55 g, 73%). Compound **4** had: ¹H NMR data (CD₃Cl₃): δ 8.05 (m, 2 H, ArH), 7.10 (t, 2 H, *J* 8.7 Hz, ArH), 4.35 (t, 2 H, *J* 6.5 Hz, CH₂), 3.73 (t, 2 H, *J* 6.4 Hz, CH₂), 1.87 (m, 2 H, CH₂), 1.80 (br s, 1 H, OH) 1.72 (m, 2 H, CH₂). ¹⁹F NMR data (CDCl₃): δ -106.2 (s, 1 F).

Resin 5

DEAD (0.538 mL, 3.42 mmol, in 1.5 mL THF) was added dropwise during 30 min to a mixture of resin **3** (0.228 mmol, prewashed with 2×8 mL THF), PPh₃ (90 mg, 3.43 mmol) and **4** (0.73 g, 3.42 mmol) in THF (1.5 mL) at -5 °C. After 3 h of vertical agitation the temperature was raised to 0 °C. After 21 h the resin was washed with THF, DMF,

20% piperidine in DMF, DMF and CH₂Cl₂ (5×5 mL each). Resin 5 had: ¹⁹F NMR data (CDCl₃): δ -106.3, -134.4 (2s each 1F).

Resin 6

Sodium methoxide in methanol (0.2 M, 1.2 mL) was added to a suspension of resin 5 (0.228 mmol) in CH₂Cl₂ (4.8 mL). After 3 h of vertical agitation the resin was washed with CH₂Cl₂, MeOH, CH₂Cl₂ containing 3% HOAc, CH₂Cl₂, DMF and CH₂Cl₂ (3×5 mL each). Resin 6 had: ¹⁹F NMR data (CDCl₃): δ -134.5 (s, 1F).

Resin 7

MSNT (0.27 g, 0.91 mmol) and Fmoc-Ser(*t*Bu)-OH (0.35 g, 0.91 mmol) were dissolved in CH₂Cl₂ (2.5 mL) and added to resin 6 (0.228 mmol). Methyl imidazole (55 µl, 0.69 mmol) was added and the mixture was agitated vertically during 4 h. The resin was washed with CH₂Cl₂, DMF and CH₂Cl₂ (6×6 mL each). The coupling was repeated once. Resin 7 had: ¹⁹F NMR data (CDCl₃): δ -134.3 (s, 1F).

Resin 8

Resin 7 (0.228 mmol) was treated with 20% piperidine in DMF (2×6 mL, 5 min vertical agitation) and washed with DMF and CH₂Cl₂ (5×6 mL each). DIC (137.6 µl, 0.89 mmol) was added to a solution of 4-fluorobenzoic acid (128 mg, 0.91 mmol) and HOBt (185 mg, 1.37 mmol) in DMF (5 mL). The mixture were transferred to the resin and Bromophenol Blue was added (0.001 equiv.) as indicator of free amino groups. After 20 h agitation the resin was washed with DMF and CH₂Cl₂ (5×6 mL each). The resin was suspended in CH₂Cl₂ (3.5 mL) and Ac₂O (1.5 mL) was added. After 5 h agitation

the resin was washed CH₂Cl₂, DMF and CH₂Cl₂ (5×6 mL each). TFA/H₂O (9:1, 5 mL) was added to the resin and after 2 h of agitation the resin was washed with HOAc (5×6 mL), CH₂Cl₂ (5×6 mL), 20% piperidine in DMF (6 mL), DMF (5×6 mL), CH₂Cl₂ (5×6 mL) and then dried under vacuum. Resin **8** had: ¹⁹F NMR data (CDCl₃): δ -108.4, -134.3 (2s each 1F).

4-Methylphenyl 2,3,4,6-tetra-*O*-(4-fluorobenzoyl)-1-thio-β-D-galactopyranoside (9**)**

Sodium methoxide in methanol (0.2 M, 2mL) was added to a solution of 4-methylphenyl 2,3,4,6-tetra-*O*-acetyl-1-thio-β-D-galactopyranoside (3.75 g, 8.25 mmol) in methanol (38 mL). After stirring for 2 h the mixture was neutralized with Amberlite-H IR-120, filtered and co-concentrated with toluene. The crude product and DMAP (0.4 g, 3.3 mmol) were dissolved in pyridine (25 mL), and 4-fluorobenzoyl chloride (5.85 mL, 49.5 mmol) was added dropwise over 10 min. The mixture was stirred for 18 h and then diluted with CH₂Cl₂ (200 mL), washed with sat. aq. NaHCO₃ (3×70 mL) and H₂O (100 mL). The combined organic phases were concentrated and residual pyridine was removed by co-evaporation with toluene. The residue was re-crystallized from EtOAc to give **9** (6.1 g, 95%) as white crystals. Compound **9** had: ¹H NMR data (CD₃Cl₃): δ 8.02 (m, 4 H, ArH), 7.89 (m, 2 H, ArH), 7.75 (m, 2 H, ArH), 7.47 (d, 2 H, *J* 8.1 Hz, ArH), 7.10 (m, 8 H, ArH), 6.91 (t, 2 H, *J* 8.7 Hz, ArH), 5.94 (d, 1 H, *J* 3.0 Hz, H-4), 5.67 (t, 1 H, *J* 9.0 Hz, H-2), 5.55 (dd, 1 H, *J* 9.9, 3.2 Hz, H-3), 4.97 (d, 1 H, *J* 9.8 Hz, H-1), 4.64 (dd, 1 H, *J* 10.8, 6.2 Hz, H-6), 4.37 (m, 2 H, H-5, H-6), 2.39 (s, 3 H, PhCH₃). ¹⁹F NMR data (CDCl₃): δ -104.5, -104.9, -105.0, -105.3 (4s each 1 F).

4-Methylphenyl 2,4-di-O-(4-fluorobenzoyl)-3,6-di-O-tert-butyldimethylsilyl-1-thio- β -D-galactopyranoside (10)

Sodium methoxide in methanol (1 M, 6.7 mL) was added to a solution of 4-methylphenyl 2,3,4,6-tetra-O-acetyl-1-thio- β -D-galactopyranoside (4.98 g, 11.0 mmol) in MeOH (84 mL) at 0 °C. The solution was stirred for 1 h 15 min before being neutralized with Amberlite-H IR-120, filtered and concentrated. The crude product was dissolved in DMF (30 mL) and *tert*-butyldimethylsilyl chloride (3.38 g, 22.5 mmol) in DMF (7 mL) was added at 0 °C. The solution was allowed to attain room temperature and after stirring for 7 h the mixture was diluted with CH₂Cl₂ (500 mL) and washed with sat. aq. NH₄Cl (500 mL). The aqueous phase was extracted with CH₂Cl₂ (2×200 mL) and the combined organic phases were dried and concentrated. The residue was flash chromatographed twice (heptane/EtOAc 5:1 and heptane/EtOAc 6:1) to give 4-methylphenyl 3,6-di-O-tert-butyldimethylsilyl-1-thio- β -D-galactopyranoside (3.04 g, 54%). 4-Methylphenyl 3,6-di-O-tert-butyldimethylsilyl-1-thio- β -D-galactopyranoside (1.02 g, 1.98 mmol) and DMAP were dissolved in pyridine and 4-fluorobenzoyl chloride (1.52 mL, 5.57 mmol) was added. The mixture was stirred for 24 h and then diluted with CH₂Cl₂ (150 mL), washed with sat. aq. NaHCO₃ (2×50 mL) and H₂O (50 mL). The organic phase was concentrated and residual pyridine was removed by evaporation with toluene. Flash chromatography of the residue (heptane/EtOAc 12:1) gave **10** (0.99 g, 79%). Compound **10** had: ¹H NMR data (CD₃Cl₃): δ 8.07 (m, 2 H, ArH), 7.94 (m, 2 H, ArH), 7.44 (d, 2 H, *J* 8.0 Hz, ArH), 7.12 (m, 6 H, ArH), 5.61 (s, 1 H, H-4), 5.42 (t, 1 H, *J* 8.9 Hz, H-2), 4.77 (d, 1 H, *J* 9.7 Hz, H-1), 4.04 (d, 1 H, *J* 8.5 Hz, H-3), 3.82 – 3.66 (m, 3 H, H-5, 2 H-6), 2.38 (s, 3 H, PhCH₃), 1.55 (s, 9 H,

*t*Bu), 1.26 (s, 9 H, *t*Bu), 0.05 (s, 3 H, SiCH₃), 0.02 (s, 3 H, SiCH₃), -0.01 (s, 3 H, SiCH₃), -0.17 (s, 3 H, SiCH₃). ¹⁹F NMR data (CDCl₃): δ -106.0, -106.3 (2s each 1 F).

4-Methylphenyl 2,3-di-*O*-(4-fluorobenzoyl)-4,6-*O*-(4-fluorobenzylidene)-1-thio-β-D-galactopyranoside (11)

Metanolic sodium methoxide (2 mM, 50 mL) was added to crystals of 4-methylphenyl 2,3,4,6-tetra-*O*-benzoyl-1-thio-β-D-galactopyranoside (7.0 g, 9.96 mmol). After stirring for 19 h the mixture was neutralized with Amberlite-H IR-120, filtered and co-concentrated with toluene. The crude product and α,α-dimethoxy-4-fluorotoluene (2.02 mL, 13.0 mmol) were dissolved in MeCN (15 mL) and *para*-toluenesulphonic acid (284 mg, 1.49 mmol) was added. After stirring for 20 h the mixture was neutralized with Et₃N (0.4 mL) and the solvent was evaporated. Flash chromatography of the residue (heptane/EtOAc 3:1) gave 4-methylphenyl 4,6-*O*-(4-fluorobenzylidene)-1-thio-β-D-galactopyranoside (3.50 g, 90%).

4-Fluorobenzoyl chloride (0.78 mL, 6.60 mmol) was added to a solution of 4-methylphenyl 4,6-*O*-(4-fluorobenzylidene)-1-thio-β-D-galactopyranoside (1.00 g, 2.55 mmol) in pyridine (10 mL). The mixture was stirred for 17 h and then diluted with CH₂Cl₂ (150 mL), washed with sat. aqueous NaHCO₃ (3×50 mL) and H₂O (100 mL). The organic phase was concentrated and residual pyridine was removed by co-evaporation with toluene. Flash chromatography of the residue (heptane/EtOAc 5:1 → 3:1) gave **11** (1.28 g, 79%). Compound **11** had: ¹H NMR data (CD₃Cl₃): δ 8.00 (m, 2 H, ArH), 7.93 (m, 2 H, ArH), 7.49 (d, 2 H, *J* 8.1 Hz, ArH), 7.35 (m, 2 H, ArH), 7.05 (m, 8 H, ArH), 5.69 (t, 1 H, *J* 9.9 Hz, H-2), 5.47 (s, 1 H, 4-FPhCH), 5.31 (dd, 1 H, *J* 10.0, 3.3 Hz, H-3), 4.87 (d, 1 H, *J* 9.8 Hz, H-1), 4.55 (d, 1 H, *J* 3.2 Hz, H-4), 4.43 (dd, 1 H, *J* 12.4, 1.2 Hz,

H-6), 4.08 (dd, 1 H, J 12.4, 1.2 Hz, H-6), 3.74 (s, 1 H, H-5), 2.36 (s, 3 H, PhCH₃). ¹⁹F NMR data (CDCl₃): δ -105.1, -105.5, -113.2 (3s each 1 F).

Resin 12, glycosylation promoted by DMTST

Dimethyl(methylthio)-sulphonium triflate (174 mg, 0.67 mmol), **9** (0.17 mmol) and resin **8** (0.057 mmol) were dried under vacuum for 4 h, after which CH₂Cl₂ (2.5 mL) was added. After 20 h of vertical agitation the resin was washed with CH₂Cl₂ (6×3 mL), 20% piperidine in DMF (3×3 mL), DMF (3×3 mL) and CH₂Cl₂ (6×3 mL). Resin **12** had: ¹⁹F NMR data (CDCl₃): δ -104.2 (s, 0.23 F), -104.3 (s, 0.16 F), -104.6 (s, 0.27 F), -104.7 (s, 0.29 F), -104.8 (s, 0.08 F), -105.1 (s, 0.31 F), -105.3 (s, 0.07 F), -107.6 (s, 0.67 F), -107.7 (s, 0.07 F), -107.8 (s, 0.25 F), -134.3 (s, 1 F).

Resin 12, glycosylation with triflic acid and *N*-iodosuccinimide (NIS)

Triflic acid (4 μ L, 46 μ mol) was added to a solution of resin **8** (0.11 mmol), NIS (128 mg, 0.57 mmol) and **9** (442 mg, 0.57 mmol) in CH₂Cl₂ (3 mL) and the mixture was agitated vertically in the absence of light. After 3 h the resin was washed with CH₂Cl₂ (5×3 mL), THF (5×3 mL), 20% piperidine in DMF (5×3 mL), DMF (5×3 mL), CH₂Cl₂ (5×3 mL) and then dried under vacuum. Resin **12** had: ¹⁹F NMR data (CDCl₃): δ -104.1, -104.5, -104.6, -105.0, -107.8, -134.5 (6s each 1 F).

Resin 14, glycosylation with triflic acid and *N*-iodosuccinimide (NIS)

Triflic acid (2.1 μ L, 24 μ mol) was added to a solution of resin **8** (0.12 mmol), NIS (134 mg, 0.60 mmol) and **9** (452 mg, 0.60 mmol) in CH₂Cl₂ (3 mL) and the mixture was agitated vertically in the absence of light at. After 3 h the resin was washed with

CH₂Cl₂ (5×3 mL), THF (5×3 mL), 20% piperidine in DMF (5×3 mL), DMF (5×3 mL) and CH₂Cl₂ (5×3 mL) and then dried under vacuum. Resin **14** had: ¹⁹F NMR data (CDCl₃): δ -105.2, -105.4, -105.9, -106.0 (4s, 2 F), -108.0 (s, 0.15 F), -108.2 (s, 0.84 F), -134.3 (s, 1 F).

***N*^α-(4-Fluorobenzoyl)-3-O-(β-D-galactopyranosyl)-L-serine (**13**)**

From resin 12: A solution of LiOH in H₂O (30 mM, 4 mL) and THF (2 mL) was added to resin **12** (40 μmol). After 3 h of vertical agitation, more LiOH in H₂O (2 M, 0.12 mL) was added. The mixture was agitated for an additional 2 h and then filtered. The resin was washed with HOAc (2×3 mL) and H₂O (2×3 mL). The combined filtrates were freeze dried and the residue was purified with reversed-phase HPLC (gradient: 100% A → 100% B during 60 min) to give **13** (6 mg, 39%). Gel-phase ¹⁹F NMR spectroscopy revealed complete cleavage of **13** from the resin.

From resin 14: Resin **14** (54 μmol) was agitated with TFA/H₂O (9:1, 4 mL) for 4 h and then washed with THF (3×3 mL), 20% piperidine in DMF (3×3 mL), DMF (5×3 mL) and CH₂Cl₂ (6×3 mL). The resin was then treated as in the cleavage of resin **12**. The crude product was purified with reversed phase HPLC (gradient: 100% A → 100% B during 60 min) to give **13** (4 mg, 19%). Gel-phase ¹⁹F NMR spectroscopy revealed complete cleavage of **13** from the resin.

Compound **13** had: ¹H NMR data (CD₃OD): δ 7.96 (m, 2 H, ArH), 7.19 (t, 2 H, *J* 6.83 Hz, ArH), 4.45 (dd, 1 H, *J* 10.4, 4.7 Hz, Ser-H_α), 4.28 (d, 1 H, *J* 7.6 Hz, H-1), 3.93 (dd, 1 H, *J* 10.4, 3.5 Hz, Ser-H_β), 3.82 (d, 1 H, *J* 2.9 Hz, H-4), 3.47 (dd, 1 H, *J* 9.8, 3.2 Hz, H-3). ¹³C NMR data (CD₃OD): δ 169.08, 167.60, 165.10, 131.71, 131.27, 116.36, 105.55, 76.94, 74.88, 72.55, 70.77, 70.35, 62.52, 55.06. ¹⁹F NMR data (CD₃OD/CDCl₃): δ -108.5 (s, 1 F).

Resin 15, glycosylation with triflic acid and *N*-iodosuccinimide (NIS)

Triflic acid (2.3 μ l, 25 μ mol) was added to a solution of resin **8** (0.17 mmol), NIS (190 mg, 0.84 mmol) and **9** (0.54 g, 0.84 mmol) in CH₂Cl₂ (3 mL) and the mixture was agitated vertically in the absence of light at ambient temperature. After 3h the resin was washed with CH₂Cl₂ (5 \times 3 mL), THF (5 \times 3 mL), 20% piperidine in DMF (5 \times 3 mL), DMF (5 \times 3 mL), CH₂Cl₂ (5 \times 3 mL) and then dried under vacuum. Resin **15** had: ¹⁹F NMR data (CDCl₃): δ -105.0, -105.3, -108.5, -113.2, -134.8 (5s, each 1 F).

***N* ^{α} -(4-Fluorobenzoyl)-3-O-[(4,6-*O*-4-fluorbenzylidene)- β -D-galactopyranosyl]-L-serine (**16**)**

Resin **15** (54 μ mol) was treated as in the cleavage of resin **12**. The residue was purified with reversed-phase HPLC (gradient: 100% *A* \rightarrow 100% *B* in 60 min) and flash chromatography (CH₂Cl₂/MeOH 10:1 \rightarrow 3:1) to give **16** (10 mg, 37%). Gel-phase ¹⁹F NMR spectroscopy revealed complete cleavage of **16** from the resin. Compound **16** had: ¹H NMR data (CD₃OD): δ 7.90 (dd, 2 H, *J* 8.7, 5.3 Hz, ArH), 7.48 (dd, 2 H, *J* 8.6, 5.5 Hz, ArH), 7.02 (m, 4 H, ArH), 5.55 (s, 1 H, 4-FPhCH), 4.43 (dd, 1 H, *J* 10.4, 4.0 Hz, Ser-H α), 4.33 (d, 1 H, *J* 6.9 Hz, H-1), 4.21 (d, 1 H, *J* 12.5 Hz, H-6), 4.16 (d, 1 H, *J* 2.7 Hz, H-4), 4.08 (dd, 1 H, *J* 12.4, 1.1 Hz, H-6), 3.92 (dd, 1 H, *J* 10.1, 2.2 Hz, Ser-H β), 3.62 (m, 2 H, H-3, H-2), 3.49 (s, 1 H, H-5). ¹³C NMR data (CD₃OD): δ 168.14, 165.63, 165.02, 163.18, 136.00, 131.76, 131.25, 129.64, 116.38, 115.65, 105.50, 101.59, 77.46, 73.55, 72.07, 71.50, 69.84, 68.14, 56.23. ¹⁹F NMR data (CD₃OD/CDCl₃): δ -108.46, -113.44 (2s each 1 F).